Biomed. Data Science: Unsupervised Datamining





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Structure of Genomic Features Matrix



Unsupervised Mining

- Simple overlaps & enriched regions
- Clustering rows & columns (networks)
- PCA/SVD (theory + appl.)
- Biplot
- RCA
- CCA
- tSNE
- LDA
- (Variational Autoencoders)

Genomic Features Matrix: Deserts & Forests



Non-random distribution of TREs

- TREs are not evenly distributed throughout the encode regions $(P < 2.2 \times 10^{-16})$.
- The actual TRE distribution is power-law.
- The null distribution is 'Poissonesque.'
- Many genomic subregions with extreme numbers of TREs.



Number of TREs in a subregion

Aggregation & Saturation

B Saturation Analysis Genome Coverage by Fraction of 2 2+3 1+3 3 1+2,3+4 2+4 1+4 all rows any any 1 row any 3 rows 2 rows C Aggregation Analysis Signal track Anchor track Λ 2 3 4

Nat. Rev. Genet. (2010) 11: 559

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Clustering Columns & Rows of the Data Matrix

Correlating Rows & Columns



[*Nat. Rev. Genet.* (2010) 11: 559]

Spectral Methods Outline & Papers

- Simple background on PCA (emphasizing lingo)
- Expression Clustering
- More abstract run through on SVD
- Application to
 - O Alter et al. (2000). "Singular value decomposition for genomewide expression data processing and modeling." PNAS 97: 10101
 - Langfelder P, Horvath S (2007) Eigengene networks for studying the relationships between co-expression modules. BMC Systems Biology 2007, 1:54
 - Z Zhang et al. (2007) "Statistical analysis of the genomic distribution and correlation of regulatory elements in the ENCODE regions." Genome Res 17: 787
 - TA Gianoulis et al. (2009) "Quantifying environmental adaptation of metabolic pathways in metagenomics." PNAS 106: 1374.

Expression Clustering



- Single or multilink
 - threshold for connection?

http://commons.wikimedia.org/wiki/File:Hierarchical_clustering_diagram.png

K-means



1) Pick ten (i.e. k?) random points as putative cluster centers.

2) Group the points to be clustered by the center to which they are closest.

3) Then take the mean of each group and repeat, with the means now at the cluster center.

4)Stop when the centers stop moving.

[Brown, Davis]

Clustering the yeast cell cycle to uncover interacting proteins





Microarray timecourse of 1 ribosomal protein

Clustering the yeast cell cycle to uncover interacting proteins





Random relationship from ~18M

Clustering the yeast cell cycle to uncover interacting proteins



[Botstein; Church, Vidal]



Close relationship from 18M (2 Interacting Ribosomal Proteins)

Clustering the yeast cell cycle to uncover interacting proteins





Global Network of Relationships



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SVD

Puts together slides prepared by Brandon Xia with images from Alter et al. papers

SVD for microarray data (Alter et al, PNAS 2000)



$A = USV^T$

- A is any rectangular matrix $(m \ge n)$
- Row space: vector subspace generated by the row vectors of A
- Column space: vector subspace generated by the column vectors of A
 - The dimension of the row & column
 space is the rank of the matrix A: r (≤ n)
- A is a linear transformation that maps vector x in row space into vector Ax in column space



Genes

$A = USV^T$

- U is an "orthogonal" matrix $(m \ge n)$
- Column vectors of U form an orthonormal basis for the column space of A: U^TU=I

$$U = \begin{pmatrix} | & | & | \\ \mathbf{u}_1 & \mathbf{u}_2 & \cdots & \mathbf{u}_n \\ | & | & | \end{pmatrix}$$

Eigenarrays



- $u_1, ..., u_n$ in U are eigenvectors of AA^T - $AA^T = USV^T VSU^T = US^2 U^T$
 - "Left singular vectors"

$A = USV^T$

- V is an orthogonal matrix (n by n)
- Column vectors of V form an orthonormal basis for the row space of A: V^TV=VV^T=I

Arrays



"Right singular vectors"

$A = U\mathbf{S}V^T$

- S is a diagonal matrix (n by n) of nonnegative singular values
- Typically sorted from largest to smallest
- Singular values are the non-negative square root of corresponding eigenvalues of A^TA and AA^T

Eigengenes

4010HO HHHHO0001001HM0H



AV = US

- Means each $Av_i = s_i u_i$
- Remember A is a linear map from row space to column space
- Here, A maps an orthonormal basis {v_i} in row space into an orthonormal basis {u_i} in column space
- Each component of u_i is the projection of a row of the data matrix A onto the vector v_i

SVD as sum of rank-1 matrices

- $A = USV^T$
- $A = s_1 \boldsymbol{u}_1 \boldsymbol{v}_1^T + s_2 \boldsymbol{u}_2 \boldsymbol{v}_2^T + \dots + s_n \boldsymbol{u}_n \boldsymbol{v}_n^T$
- an outer product (uv^T) giving a matrix rather than the scalar of the inner product
- What is the rank-r matrix that best approximates A ?

- Minimize
$$\sum_{i=1}^{m} \sum_{j=1}^{n} (\hat{A}_{ij} - A_{ij})^2$$

• $s_1 \ge s_2 \ge \ldots \ge s_n \ge 0$

LSQ approx. If r=1, this amounts to a line fit.

- $\hat{A} = s_1 \boldsymbol{u}_1 \boldsymbol{v}_1^T + s_2 \boldsymbol{u}_2 \boldsymbol{v}_2^T + \dots + s_r \boldsymbol{u}_r \boldsymbol{v}_r^T$
- Very useful for matrix approximation

Examples of (almost) rank-1 matrices

- Steady states with fluctuations
- Array artifacts?
 101 303 202
 102 300 201
 103 304 203
 101 302 204
- Signals?

$$\begin{pmatrix} 1 & 2 & -1 \\ 2 & 4 & -2 \\ -1 & -2 & 1 \\ 0 & 0 & 0 \end{pmatrix}$$

101	103	102
302	300	301
203	204	203
401	402	404)

Geometry of SVD in row space



This line segment that goes through origin approximates the original data set

The projected data set approximates the original data set

Geometry of SVD in row space

- A as a collection of m row vectors (points) in the row space of A
- $s_1 u_1 v_1^T + s_2 u_2 v_2^T$ is the best rank-2 matrix approximation for A
- Geometrically: v₁ and v₂ are the directions of the best approximating rank-2 subspace that goes through origin
- s₁u₁ and s₂u₂ gives coordinates for row vectors in rank-2 subspace
- *v*₁ and *v*₂ gives coordinates for row space basis vectors in rank-2 subspace



$$A\mathbf{v}_{\mathbf{i}} = s_{i}\mathbf{u}_{\mathbf{i}}$$

$$I \mathbf{v}_{\mathbf{i}} = \mathbf{v}_{\mathbf{i}}$$
 28

What about geometry of SVD in column space?

- $A = USV^T$
- $A^T = VSU^T$
- The column space of A becomes the row space of A^T
- The same as before, except that U and V are switched

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Intuition on interpretation of SVD in terms of genes and conditions

Genes sorted by correlation with top 2 eigengenes



Alter, Orly et al. (2000) Proc. Natl. Acad. Sci. USA 97, 10101-10106

Fig. 3. Genes sorted by relative correlation with $|\gamma_1\rangle_N$ and $|\gamma_2\rangle_N$ of normalized elutriation. (a) Normalized elutriation expression of the sorted 5,981 genes in the 14 arrays, showing traveling wave of expression. (b) Eigenarrays expression; the expression of $|\alpha_1\rangle_N$ and $|\alpha_2\rangle_N$, the eigenarrays corresponding to $|\gamma_1\rangle_N$ and $|\gamma_2\rangle_N$, displays the sorting. (c) Expression levels of $|\alpha_1\rangle_N$ (red) and $|\alpha_2\rangle_N$ (green) fit normalized sine and cosine functions of period $Z \equiv N - 1 = 5,980$ and phase $\theta \approx 2\pi/13$ (blue), respectively.



Normalized elutriation expression in the subspace associated with the cell cycle



Alter, Orly et al. (2000) Proc. Natl. Acad. Sci. USA 97, 10101-10106



Fig. 2. Normalized elutriation expression in the subspace associated with the cell cycle. (a) Array correlation with $|\alpha_1\rangle_N$ along the *y*-axis vs. that with $|\alpha_2\rangle_N$ along the *x*-axis, color-coded according to the classification of the arrays into the five cell cycle stages, M/G₁ (yellow), G₁ (green), S (blue), S/G₂ (red), and G₂/M (orange). The dashed unit and half-unit circles outline 100% and 25% of overall normalized array expression in the $|\alpha_1\rangle_N$ and $|\alpha_2\rangle_N$ subspace. (b) Correlation of each gene with $|\gamma_1\rangle_N$ vs. that with $|\gamma_2\rangle_N$, for 784 cell cycle regulated genes, color-coded according to the classification by Spellman *et al.* (3).

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Biplot

Introduction



- A biplot is a lowdimensional (usually 2D) representation of a data matrix A.
 - A point for each of the *m* observation vectors (rows of A)
 - A line (or arrow) for each of the *n* variables (columns of A)

ΔΤ

а

b

С

з

4 5 6 - 7 8

16 18 17 19 23 14 21 18 13 10

28 25 22 33 28 34 30 22 36 32

2



AA^T (site-site correlation)

0.97 0.50 0.49 0.59 0.31 0.93 0.84 0.43 0.89 1.00

10



5

2

0.5

-0.5

0.0

Principal component V1

8

з

1.0

PCA

TFs: a, b, c Genomic Sites: 1,2,3	Biplot to Show Ove TFs &	erall Relationship of Sites
A=USV a b c 1 21 16 28 1 4 18 25 3 14 17 22 4 14 19 33 5 17 23 28 6 20 14 34 7 22 21 30 8 15 18 22 9 18 13 36 10 24 10 32	a b c a 1.00 -0.44 0.48 -0.44 1.00 -0.40 0.48 -0.40 1.00 A ^T A (TF-TF corr.)	$Lincipal component VI^{Z}$
A T 1 2 3 4 5 6 7 a 21 14 14 14 17 20 22 1 b 16 18 17 19 23 14 21 1 c 28 25 22 33 28 34 30 2	1 2 3 4 5 6 7 8 9 1 2 3 4 5 6 7 8 9 1 <th>$\begin{array}{c} 10 \\ 0.97 \\ 0.59 \\ 0.49 \\ 0.43 \\ 0.43 \\ 0.43 \\ 0.43 \\ 0.43 \\ 0.64 \\ 0.6$</th>	$ \begin{array}{c} 10 \\ 0.97 \\ 0.59 \\ 0.49 \\ 0.43 \\ 0.43 \\ 0.43 \\ 0.43 \\ 0.43 \\ 0.64 \\ 0.6$

0.5 -1.0 -0.5 0.0 1.0 Principal component U1



Results of Biplot

Zhang et al. (2007) Gen. Res.

- Pilot ENCODE (1% genome): 5996 10 kb genomic bins (adding all hits) + 105 TF experiments \rightarrow biplot
- Angle between TF vectors shows relation b/w factors
- Closeness of points gives clustering of "sites"
- Projection of site onto vector gives degree to which site is assoc. with a particular factor



Results of Biplot

Zhang et al. (2007) Gen. Res.

- Biplot groups TFs into sequence-specific and sequence-nonspecific clusters.
 - c-Myc may behave more like a sequence-nonspecific TF.
 - H3K27me3 functions in a transcriptional regulatory process in a rather sequence-specific manner.
- Genomic Bins are associated with different TFs and in this fashion each bin is "annotated" by closest TF cluster

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RCA

What is RCA?

- RCA stands for **Reference** Component Analysis
- RCA is an algorithm that expands the standard PCA to address noisy data:
 - Batch effect
 - Low signal to noise datasets
- It is still an unsupervised clustering method but, RCA adds external information to address noisy data:
 - Instead of projecting the original data into new axis
 - It first correlates the original data to a reference panel
 - And then, performs PCA on the correlations
- In single-cell or bulk RNA-seq

Projection to external dataset



Correlation matrix





S2

S1

 $C = U * V^T$

PC1⁵

Ó

10

-5

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CCA

Sorcerer II Global Ocean Survey



Sorcerer II Global Ocean Survey





Expressing data as matrices indexed by site, env. var., and pathway usage

[Rusch et. al., (2007) PLOS Biology; Gianoulis et al., PNAS (in press, 2009]



Canonical Correlation Analysis: Simultaneous weighting





Canonical Correlation Analysis: Simultaneous weighting



CCA: Finding Variables with Large Projections in "Correlation Circle"



The goal of this technique is to interpret cross-variance matrices We do this by defining a change of basis.

Gianoulis et al., PNAS 2009



Strength of Pathway co-variation with environment



Environmentally Environmentally invariant variant



Gianoulis et al., PNAS 2009

Conclusion #1: energy conversion strategy, temp and depth



Gianoulis et al., PNAS 2009

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tSNE

tSNE

a technique for dimensionality reduction that is particularly well suited for the visualization of high-dimensional datasets



hyperparameters 'perplexity' really matter; Cluster sizes in a t-SNE plot mean nothing; Distances between clusters might not mean anything

https://distill.pub/2016/misread-tsne/

https://scikit-learn.org/stable/auto_examples/manifold/plot_t_sne_perplexity.html#sphx-glr-auto-examples-manifold-plot-t-sne-perplexity-py

Example: t-SNE clustering of **14,685** single-cell transcriptomes



Comparison on real datasets (melanoma scRNA-seq dataset)



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LDA









https://towardsdatascience.com/light-on-math-machine-learning-intuitive-guide-to-latent-dirichlet-allocation-437c81220158

Diagram



```
oldsymbol{arphi}_{k=1\ldots K} \sim \mathrm{Dirichlet}_V(oldsymbol{eta}) \ oldsymbol{	heta}_{d=1\ldots M} \sim \mathrm{Dirichlet}_K(oldsymbol{lpha}) \ z_{d=1\ldots M,w=1\ldots N_d} \sim \mathrm{Categorical}_K(oldsymbol{	heta}_d) \ w_{d=1\ldots M,w=1\ldots N_d} \sim \mathrm{Categorical}_V(oldsymbol{arphi}_{z_{dw}})
```

 $\boldsymbol{\alpha}$ is the parameter of the Dirichlet prior on the per-document topic distributions

 β is the parameter of the Dirichlet prior on the per-topic word distribution

 θ_i is the topic distribution for document i

 φ_k is the word distribution for topic k

 z_{ij} is the topic for the jth word in document i

 w_{ij} is the specific word, and

K is the number of topics, N is the number of word in a document, M is the number of Documents.



The sparsity is important

hyperparameter of Dirichlet distribution enable the sparsity of document to topic (θ) and word to topic (φ) distribution, make LDA works better than others similar methods most of time.



In LDA analysis, alpha should be tuned for topic distribution(**θ**)

How the distribution of θ changes with different α values

https://towardsdatascience.com/light-on-math-machinelearning-intuitive-guide-to-latent-dirichlet-allocation-437c81220158

Comparison of sparsity using **USArrests** dataset in a three-dimensional space



PCA Rotation=XW(loading)





NMF (rank=3) X=W*H W

